triple-drug therapy does not seem to select significant escape mutation, unlike single- or double-drug therapy; part of the protection must be due to the drug-induced low level of virus replication. This gives some hope that if the immune response is activated very early in infection, there is a chance that the virus could be controlled by a response to at least three epitopes before high-level virus replication occurs. Given the above arguments about pre-existing virus variability, this could mean inducing CTL responses to ten or more epitopes.

Breadth of the immune response

A remarkable feature of the natural T-cell response to acute or chronic virus infection is that the CD8+ T-cell response can be focused on a very small number of epitopes^{77,78}. In the CD8+ T-cell response to acute EBV infection, as many as 40% of blood CD8+ T cells can respond to a single epitope13, despite the fact that this herpes virus expresses hundreds of proteins. This type of CTL response could be disastrous for a vaccine, as it offers an easy escape route. It is not clear how to broaden a vaccine response, and the obvious possibility of adding more virus proteins to the vaccine might not work (as for EBV). It might be better to mix several small vaccine constructs together, fooling the immune system into responding to several 'invaders', each requiring a strong T-cell response. For a DNA prime and recombinant virus boost schedule, it might only be necessary to do this for the DNA priming component.

Duration of the immune response

In macaques that were immunized with nonreplicating MVA, the half life of tetramerstained CD8+ T cells seems to be around seven days⁶⁸. The memory T-cell response that remains is at a much lower level. This is probably typical of the response to a non-persisting antigen. If a high level of mature effectors is required for protection, continuous or repeated antigenic stimulation will be required. The evidence from the Nairobi sex workers indicates that this will be needed, at least for complete protection; in several cases. susceptibility to HIV infection was restored when they ceased prostitution 109. However, their concentration of antigen-specific CD8+ T cells while they were protected was less than that which can be induced in humans by a vaccine, so the situation might not be exactly comparable. Amara et al.64 found that their macaques were protected against SHIV-89.6P disease when challenged seven months after the last immunization. By contrast, there was little or no protection in macaques challenged

at the peak of the tetramer response to a single epitope⁶⁸ (T. M. Allen, T. Hanke and D. Watkins, unpublished observations).

The SHIV-89.6P-challenge studies indicate that useful but incomplete protection can be obtained by long-lasting memory T cells; complete protection might need higher levels of fully activated effectors. Only phase III efficacy trials will show whether CTL memory that is induced by current vaccines will work. If non-persisting vaccines do not protect, persisting antigen vaccines will have to be tested. The regulatory authorities will have to confront this need.

Why the idea might be right but fail

The animal studies that have been discussed show that the CTL-vaccine approach can work. However, for HIV, conditions will have to be exactly right. There is a danger that one or two negative trials could kill the whole idea of a CTL vaccine. Therefore, it is vital that conditions for the first efficacy trials are optimal. The reasons that a vaccine might fail have been discussed: the vaccine-induced T cells might have to be in an activated state that cannot be maintained by the vaccine, the immune response might be too weak, the T cells might not see enough epitopes to cope with virus variability, the virus might escape from the T-cell response or the duration of protection might only be brief. Even at best, a CTL-inducing vaccine might be only half a vaccine — that is, it might only really protect in combination with neutralizing antibody.

These concerns combine to produce a formidable challenge, but one that cannot be avoided. There is now a CTL-vaccine bandwagon, with several teams gearing up to test the same hypothesis. One bad trial could ruin

Glossary

CLADE

A subgroup of HIV variants with a greater degree of genome homology.

LICENSING

The activation of dendritic cells by CD4° T cells through CD40-CD40L interaction.

PRIME-BOOST

When a single application of a vaccine is insufficient, repeated immunizations are performed using the same vaccine preparation (homologous prime boost) or using different vaccine preparations (heterologous prime boost) to sequentially stimulate a better immune response.

TETRAMEI

A reagent composed of four MHC-peptide complexes linked by biotin and streptavidin, which can be fluorescently labelled and used to track antigen-specific T cells by flow cytometry.

the whole lot, although a good trial that gives a clear negative answer would be scientifically very important. A positive protective effect would open the door to a vaccine and could be within our grasp soon.

Trials: ethical and political issues

The need for an HIV vaccine is desperate in developing countries. Apart from a few exceptional sites, only these countries have a high enough incidence of HIV infection to conduct phase III efficacy trials. Therefore, it is essential to establish strong collaborations well in advance of such trials. Matching of clades between vaccine and the most prevalent virus has been used as a political argument to ensure that such collaborations are truly in the interests of the African or Asian partner. In fact, there are stronger scientific arguments as to why the clades should be matched in any phase III efficacy trial. It is also important to have an outline plan for further vaccine development to ensure that, if the vaccine works, it will be made available in the partner country at the earliest opportunity. The medical, scientific and regulatory authorities are well aware of these issues, and trials need at least two years of preparation to deal with these issues before the trial itself. The associated infrastructure development needs similar forward planning.

Thought also has to be given to serious ethical issues. The trials will only give answers if some control volunteers become infected with HIV. If the vaccine only partly protects, or does not work, vaccine recipients will also be infected. The level of treatment that they should be offered - for life - needs very careful discussion, which must involve the community to which the trial participant belongs. For these and more commercial reasons, vaccines that are targeted at developing countries are not attractive to the pharmaceutical industry. Alternative funding streams have been created and need continued support (for further discussion of these issues, see http://www.iavi.org).

Conclusions

HIV presents an unprecedented challenge to vaccine design and conduct of trials. Virus variability is a particularly serious problem. It must not be assumed that 90% similarity between HIV clades means that a vaccine that is based on one clade will give 90% protection against another clade; it is more likely that such cross protection would be as little as 33%. Efforts must be made, therefore, to ensure that the vaccine stimulates a broad response. This is difficult to ensure, given the tendency of the immune response to focus on only a few

PERSPECTIVES

epitopes — sometimes only one. These issues present formidable challenges, but if these problems are properly addressed, the animal models indicate that the vaccine will work.

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- Emini, E. A. et al. Prevention of HIV-1 infection in chimpanzees by gp120 V3 domain-specific monocional antibody. Nature 385, 728–730 (1992).
- Silvera, P. et al. Fine analysis of humoral antibody response to envelope glycoprotein of SIV in infected and vaccinated macaques. AIDS Res. Hum. Retroviruses 10, 1295–1304 (1994).
- Connor, R. I. et al. Immunological and virological analyses of persons infected by human immunodeficiency virus type 1 while participating in trials of recombinant gp120 subunit vaccines. J. Virol. 12, 1552–1576 (1998).
- Kwong, P. D. et al. Structure of an HIV gp120 envelope glycoprotein in complex with the CD4 receptor and a
- neutralizing human antibody. *Nature* **363**, 638–659 (1998).

 5. Wyatt, R. *et al.* The antigenic structure of the HIV gp120 envelope glycoprotein. *Nature* **393**, 705–711 (1998).
- Gauduin, M. C. et al. Passive immunization with a human monoclonal antibody protects hu-PBL-SCID mice against challenge by primary isolates of HIV-1. Nature Med. 3, 1389–1393 (1997)
- Med. 3, 1389–1393 (1997).
 Moore, J. P. & Burton, D. R. HIV-1 neutralizing antibodies: how full is the bottle? Nature Med. 6, 149–144 (1990).
- how full is the bottle? Nature Med. 5, 142–144 (1999).
 Polgnard, P., Saphire, E. O., Parren, P. W. & Burton, D. R. gp120: biologic aspects of structural features. Annu. Rev. Immunol. 19, 253–274 (2001).
 Daniel, M. D., Kirchhoff, F., Czajak, S. C., Sehgal, P. K. &
- Daniel, M. D., Kirchhoff, F., Czejak, S. C., Sengal, P. K. 8 Desrosiers, R. C. Protective effects of a live attenuated SIV vaccine with a deletion in the nef gene. Science 258, 1938–1941 (1992).
- Altman, J. D. et al. Phenotypic analysis of antigenspecific Tlymphocytes. Science 274, 94-96 (1996)
- Lalvani, A. et al. Rapid effector function in CO8* memory T cells. J. Exp. Med. 186, 859–865 (1997).
- Pitcher, C. J. et al. HIV-1-specific CD4* T cells are detectable in most individuals with active HIV-1 infection, but decline with prolonged viral suppression. Nature Med. 5, 518–525 (1999).
- Callan, M. F. et al. Large clonal expansions of CD8^o T cells in acute infectious mononucleosis. *Nature Med.* 2, 906–911 (1996).
- Callan, M. F. et al. CD8* T-cell selection, function, and death in the primary immune response in vivo. J. Clin. Invest 106, 1251–1261 (2000).
- Invest. 106, 1251–1261 (2000).

 15. Ogg, G. S. et al. Quantitation of HIV-1-specific cytotoxic Tymphocytes and plasma viral RNA load. Science 279, 2103–2106 (1998).
- Tan, L. C. et al. A re-evaluation of the frequency of CD6* T cells specific for EBV in healthy virus carriers. J. Immunol. 162, 1827–1835 (1999).
- Appay, V. et al. HiV-specific CD8* T cells produce antiviral cytokines but are impaired in cytolytic function. J. Exp. Med. 192, 63–75 (2000).
- Charmpagne, P. et al. Skewed maturation of memory HIVspecific CD8 T lymphocytes. Nature 410, 106–111 (2001).
- specific CD8 T lymphocytes. *Nature* 410, 106–111 (2001).

 19. Zinkernagel, R. M. The role of antigen in maintaining T cell memory. *Dev. Biol. Stand.* 82, 189–191 (1994).

 20. Lau, L. L., Jamileson, B. D., Somasundaram, T. &
- Lau, L. L., Jamieson, B. D., Somasundaram, T. & Ahmed, R. Cytotoxic T-cell memory without antigen. Nature 369, 648–652 (1994).
- Tough, D. F., Sun, S., Zhang, X. & Sprent, J. Stimulation of memory T cells by cytokines. Vaccine 18, 1642–1648 (2000).
- Oehen, S., Waldner, H., Kundig, T. M., Hengartner, H. & Zinkernagel, R. M. Antivirally protective cytotoxic T cell memory to lymphocytic choriomeningitis virus is governed by persisting antigen. J. Exp. Med. 176, 1273–1281 (1992).
- Jamieson, B. D. & Ahmed, R. T cell memory. Long-term persistence of virus-specific cytotoxic T cells. J. Exp. Med. 189, 1993–2005 (1989).
- Kalarms, S. A. & Walker, B. D. The critical need for CD4 help in maintaining effective cytotoxic T lymphocyte responses. J. Exp. Med. 188, 2199–2204 (1998).
 Zajec, A. J. et al. Viral immune evasion due to
- Zajac, A. J. et al. Viral immune evasion due to persistence of activated T cells without effector function J. Exp. Med. 188, 2205–2213 (1998).

- Shirai, M. et al. Helper-cytotoxic T lymphocyte (CTL) determinant linkage required for priming of anti-HIV CD8: CTL in vivo with peptide vaccine constructs. J. Immunol. 152, 549–556 (1994).
- Ahlers, J. D., Belyakov, I. M., Thomas, E. K. & Berzofsky, J. A. High-affinity T helper epitope induces complementary helper and APC polarization, increased CTL, and protection against viral infection. *J. Clin. Invest.* 108, 1677–1685 (2001).
- Ridge, J. P., Di Rosa, F. & Matzinger, P. A conditioned dendritic cell can be a temporal bridge between a CD4* T-helper and a T-killer cell. Nature 393, 474–478 (1999).
- Schoenberger, S. P., Toes, R. E., van der Voort, E. I., Offringa, R. & Melief, C. J. Toelf help for cytotoxic T lymphocytes is mediated by CD40-CD40L interactions. Nature 363, 480-483 (1998).
- Whitmire, J. K. et al. CD40-CD40 ligand costimulation is required for generating antitwal CD4 T cell responses but is dispensable for CD8 T cell responses. *J. Immunol.* 163, 3194–3201 (1999).
- Bennett, S. R. et al. Help for cytotoxic-T-cell responses is mediated by CD40 signaling. Nature 393, 478–480 (1998).
 Mattoubian, M., Concepcion, R. J. & Ahmed, R. CD4*
- Matloubian, M., Concepcion, R. J. & Ahmed, R. CD4⁺ T cells are required to sustain CD6⁺ cytotoxic T-cell responses during chronic viral infection. J. Virol. 68, 8056–8063 (1994).
- Walter, E. A. et al. Reconstitution of cellular immunity against cytomegalovirus in recipients of allogeneic bone marrow by transfer of T-cell clones from the donor. N. Engl. J. Med. 333, 1038–1044 (1995).
 Miskovsky, E. P. et al. Studies of the mechanism of
- Miskovsky, E. P. et al. Studies of the mechanism of cytolysis by HIV-1 -specific CD4⁺ human CTL clones induced by candidate AIDS vaccines. J. Immunol. 153, 2787–2799 (1994).
- McMichael, A. T cell responses and viral escape. Cell 93, 673–676 (1998).
- Rosenberg, E. S. et al. Vigorous HIV-1-specific CD4* T cell responses associated with control of viremia. Science 278, 1447–1450 (1997).
- Science 278, 1447-1450 (1997).

 37. Walker, B. D. et al. HIV-specific cytotoxic T lymphocytes in seropositive individuals. *Nature* 328, 345-348 (1987).
- Goulder, P. J. et al. Functionally Inert HIV-specific cytotoxic Tlymphocytes do not play a major role in chronically infected adults and children. J. Exp. Med. 192, 1819–1832 (2000).
- Koup, R. A. et al. Temporal association of cellular immune responses with the initial control of virenia in primary human immunodeficiency virus type 1 syndrome. J. Virol. 68, 4650–4655 (1994).
- Borrow, P., Lewicki, H., Hahn, B. H., Shaw, G. M. & Oldstone, M. B. Virus-specific CD8' cytotoxic T-lymphocyte activity associated with control of viremia in primary human immunodeficiency virus type 1 infection. J. Virol. 68, 6103–6110 (1994).
- Wilson, J. D. et al. Direct visualization of HIV-1-specific cytotoxic T lymphocytes during primary infection. AIDS 14, 225–233 (2000).
- McMichael, A. J. et al. The dynamics of the cellular immune response to HIV Infection: implications for vaccination. Phil. Trans. R. Soc. Lond. B Biol. Sci. 355, 1007–1011 (2000).
- Betts, M. R. et al. Analysis of total human immunodeficiency virus (HIV)-specific CD4+ and CD8+ T-cell responses: relationship to viral load in untreated HIV infection. J. Virol. 78, 11983–11991 (2001).
- Matano, T. et al. Administration of an anti-CD8 monoclonal antibody interferes with the clearance of chimeric similar/human immunodeficiency virus during primary infections of rhesus macaques. J. Virol. 72, 164–169 (1998).
- Schmitz, J. E. et al. Control of viremia in simian immunodeficiency virus infection by CD8* lymphocytes. Science 283, 857–860 (1999).
- Jin, X. et al. Dramatic rise in plasma viremia after CD8⁺ T cell depletion in simian immunodeficiency virusinfected macaques. J. Exp. Med. 188, 991–998 (1999)
- infected macaques. J. Exp. Med. 188, 991–998 (1999).
 Lifson, J. D. et al. Role of CD8: lymphocytes in control of simian immunodeficiency virus infection and resistance to rechallenge after transient early antiretroviral treatment. J. Virol. 78, 10187–10199 (2001).
- Koenig, S. et al. Transfer of HIV-1 specific cytotoxic T lymphocytes to an AIDS patient leads to selection for mutant HIV variants and subsequent disease progression. Nature Med. 1, 330–336 (1995).
- Phillips, R. E. et al. Human immunodeficiency virus genetic variation that can escape cytotoxic T cell recognition. Nature 354, 453–459 (1991).
- Goulder, P. J. et al. Late escape from an immunodominant cytotoxic T-lymphocyte response associated with progression to AIDS. Nature Med. 3, 212–217 (1997).
- progression to AIDS. Nature Med. 3, 212–217 (1997).
 Borrow, P. et al. Antiviral pressure exerted by HIV-1-specific cytotoxic T lymphocytes (CTLs) during primary infection demonstrated by rapid selection of CTL escape virus. Nature Med. 3, 205–211 (1997).

- Evans, D. T. et al. Virus-specific cytotoxic T-lymphocyte responses select for amino-acid variation in simian immunodeficiency virus Env and Nef. Nature Med. 5, 1270–1276 (1999).
- Le Gall, S. et al. Nef interacts with the μ subunit of clathrin adaptor complexes and reveals a cryptic sorting signal in MHC I molecules. *Immunity* 8, 483–495 (1998).
- Cottins, K. L., Chen, B. K., Kalams, S. A., Walker, B. D. & Baltimore, D. HIV-1 Nef protein protects infected primary cells against killing by cytotoxic T lymphocytes. *Nature* 361, 397–401 (1998).
- Rowland-Jones, S. L. et al. Cytotoxic T cell responses to multiple conserved HIV epitopes in HIV- resistant prostitutes in Nairobi. J. Clin. Invest. 102, 1758–1765 (1998).
- Dorrell, L. et al. Absence of specific mucosal antibody responses in HIV-exposed uninfected sex workers from the Gambia. AIDS 14, 1117–1122 (2000).
- Kaul, R. et al. CD8* lymphocytes respond to different HIV epitopes in seronegative and infected subjects, J. Clin. Invest. 107, 1303–1310 (2001).
- Rowland-Jones, S. L. et al. HIV-specific cytotoxic T-cell activity in an HIV-exposed but uninfected infant. Lancet 341, 860–861 (1993).
- Semand, N. F., Yannakis, C. M., Lee, J. S. & Tsoukas, C. M. Human immunodeficiency virus (HIV)-specific cytotoxic T lymphocyte activity in HIV-exposed seronegative persons. *J. Infect. Dis.* 179, 538–547 (1999).
 Pinto, L. A. *et al.* ENV-specific cytotoxic T lymphocyte
- Pinto, L. A. et al. ENV-specific cytotoxic T lymphocyte responses in HIV seronegative health care workers occupationally exposed to HIV-contaminated body fluids. J. Clin. Invest. 96, 867–876 (1995).
- Bienzle, D. et al. Factors contributing to the lack of human immunodeficiency virus type 1 (HIV-1) transmission in HIV-1-discordant partners. J. Infect. Dis. 182, 123–132 (2000).
- Lifson, J. D. et al. Containment of simian immunodeficiency virus infection: cellular immune responses and protection from rechallenge following transient postinoculation antiretroviral treatment. J. Virol. 74, 2584–2593 (2000).
- Barouch, D. H. et al. Control of viremia and prevention of clinical AIDS in rhesus monkeys by cytokine-augmented DNA vaccination. Science 290, 486–492 (2000).
- Amara, R. R. et al. Control of a mucosal challenge and prevention of AIDS by a multiprotein DNA/MVA vaccine. Science 292, 69–74 (2001).
- Rose, N. F. et al. An effective AIDS vaccine based on live attenuated vesicular stornatitis virus recombinants. Cell 106, 539–549 (2001).
- Shiver, J. W. et al. Replication-incompetent adenoviral vaccine vector elicits effective anti-immunodeficiencyvirus immunity. Nature 415, 331–335 (2002).
- Barouch, D. H. et al. Eventual AIDS vaccine failure in a rhesus monkey by viral escape from cytotoxic T lymphocytes. *Nature* 415, 335–339 (2002).
 Hanke, T. et al. Effective Induction of simian
- Hanke, T. et al. Effective Induction of simian immunodeficiency virus-specific cytotoxic T lymphocytes in macaques by using a multiepitope gene and DNA prime-modified vaccinia virus Ankara boost vaccination regimen. J. Virol. 73, 7524–7532 (1999).
- Smith, S. M. et al. Retrospective analysis of viral load and SIV antibody responses in thesus macaques infected with pathogenic SIV: predictive value for disease progression. AIDS Res. Hum. Retroviruses 15, 1691–1701 (1999).
- Webster, R. G. & Askonas, B. A. Cross-protection and cross-reactive cytotoxic T cells induced by influenza virus vaccines in mice. Eur. J. Immunol. 10, 396–401 (1980).
- McMichael, A. J., Gotch, F., Cutlen, P., Askonas, B. & Webster, R. G. The human cytotoxic T cell response to influenza A vaccination. Clin. Exp. Immunol. 43, 276–284 (1981)
- Mongkolsapaya, J. et al. Antigen-specific expansion of cytotoxic T lymphocytes in acute measles virus infection. J. Virol. 73, 67–71 (1999).
- Layton, G. T. et al. Induction of HIV-specific cytotoxic Tlymphocytes in vivo with hybrid HIV-1 V3:Ty-virus-like particles. J. Immunol. 151, 1097–1107 (1993).
 Mills, K. H. et al. Vaccine-induced CD4+T cells against
- Mills, K. H. et al. Vaccine-Induced CD4+ T cells against the similan immunodeficiency virus gag protein. Epitope specificity and relevance to protective immunity. J. Immunol. 147, 3560–3567 (1991).
- 75. Weber, J. et al. Immunogenicity of the yeast recombinant p17/p24:Ty virus-like particles (p24-VLP) in healthy volunteers. Veccine 13, 831–834 (1995).
 76. Aichele, P., Brduscha-Riem, K., Zinkernagel, R. M.,
- Archele, P., Brduscha-Riem, K., Zinkernagel, R. M., Hengartner, H. & Pircher, H. T cell priming versus T cell tolerance induced by synthetic peptides. J. Exp. Med. 182, 261–266 (1995).
- Chen, W., Anton, L. C., Bennink, J. R. & Yewdell, J. W. Dissecting the multifactonal causes of immunodominance in class I-restricted T cell responses to viruses. *Immunity* 12, 83–93 (2000).

- Yewdell, J. W. & Bennink, J. R. Immunodominance in major histocompatibility complex class I-restricted Tlymphocyte responses. *Annu. Rev. Immunol.* 17, 51–88 (1990)
- Ulmer, J. B. et al. Heterologous protection against influenza by injection of DNA encoding a viral protein. Science 259, 1745–1749 (1993).
- Science 259, 1745–1749 (1993).
 Hanke, T. et al. Enhancement of MHC class I-restricted peptide-specific T cell induction by a DNA prima/MVA boost vaccination regime. Vaccine 16, 439–445 (1998).
- Schneider, J. et al. Enhanced immunogenicity for CD8⁺ T cell induction and complete protective efficacy of malaria DNA vaccination by boosting with modified vaccinia virus Ankara. Nature Med. 4, 397–402 (1998).
- Le, T. P. et al. Safety, tolerability and humoral immune responses after intramusculer administration of a melaria DNA vaccine to healthy adult volunteers. Vaccine 18, 1833–1901 (2000).
- Wang, R. et al. Induction of antigen-specific cytotoxic T lymphocytes in humans by a malaria DNA vaccine. Science 282, 476–480 (1998).
 MacGregor, R. R. et al. First human trial of a DNA-based
- MacGregor, R. R. et al. First human trial of a DNA-based vaccine for treatment of human immunodeficiency virus type 1 infection: safety and host response. J. Infect. Dis. 178, 92–100 (1998).
- MacGregor, R. R., Boyer, J. D., Ciccerelli, R. B., Ginsberg, R. S. & Weiner, D. B. Safety and immune responses to a DNA-based human immunodeficiency virus (HIV) type I env/rev vaccine in HIV-Infected
- recipients: follow-up data: *J. Infect. Dis.* **181**, 406 (2000). 86. Ugen, K. E. *et al.* DNA vaccination with HN-1 expressing constructs elicits immune responses in humans. *Vaccine* **16**, 1818–1821 (1998).
- Boyer, J. D. et al. Vaccination of seronegative volunteers with a human immunodeficiency virus type 1 env/rev DNA vaccine induces antigen-specific proliferation and lymphocyte production of β-chemokines. J. Infect. Dis. 181, 476–483 (2000).
 Calarota, S. et al. Cellular cytotoxic response induced by
- Calarota, S. et al. Cellular cytotoxic response induced by DNA vaccination in HIV-1-infected petients. Lancet 361, 1320–1325 (1998).
- Calarota, S. A. et al. Immune responses in asymptomatic HIV-1-infected patients after HIV-DNA immunization followed by highly active antiretroviral treatment. J. Immunol. 163, 2330–2338 (1999).
- J. Immunol. 183, 2330–2338 (1999).

 90. Yewdell, J. W., Bennink, J. R., Smith, G. L. & Moss, B. Influenza A virus nucleoprotein is a major target antigen for cross-reactive anti-influenza A virus cytotoxic T lymphocytes. Proc. Natl Acad. Sci. USA 82, 1785–1789 (1985).
- Bennink, J. R., Yewdell, J. W., Smith, G. L., Moller, C. & Moss, B. Recombinant vaccinia virus primes and stimulates influenza haemagglutnini-specific cytotoxic T cells. Nature 311, 578–579 (1984).
- Clements-Mann, M. L. et al. Immune responses to human immunodeficiency virus (HIV) type 1 induced by canerypox expressing HIV-1MN gp120, HIV-1SF2 recombinant gp120, or both vaccines in seronegative adults. NIAID AIDS Vaccine Evaluation Group. J. Infect. Dis. 177, 1230–1246 (1998).
- Egan, M. A. et al. Induction of human immunodeficiency virus type 1 (HIV-1)-specific cytolytic T lymphocyte responses in seronegative adults by a nonreplicating, hostrange-restricted can any pox vector (ALVAC) carrying the HIV-1MN envigens. J. Infact. Dis. 171, 1623-1627 (1995).
 Graham, B. S. et al. Vaccination of vaccinia-naive adults
- Graham, B. S. et al. Veccination of vaccinia-naive adults with human immunodeficiency virus type 1 gp 160 recombinant vaccinia virus in a blinded, controlled, randomized clinical trial. The AIDS Vaccine Cinical Trials Network. J. Infect. Dis. 166, 244-252 (1992).
- Kantakamalakul, W. et al. Cytotoxic T lymphocyte responses to vaccinia virus antigens but not HIV-1 subtype E envelope protein seen in HIV-1 seronegative Thais. Asian Pac. J. Allergy Immunol. 19, 17–22 (2001).
 Zagury, D. et al. A group specific anamnestic immune
- zagury, J. *et al.* A group specinic anaminestic immune reaction against HIV-1 induced by a candidate vaccine against AIDS. *Nature* 332, 728–731 (1988).
 Camuth, L. M. *et al.* An algorithm for evaluating human
- Carruth, L. M. et al. An algorithm for evaluating human cytotoxic T lymphocyte responses to candidate AIDS vaccines. AIDS Res. Hum. Retroviruses 15, 1021–1034 (1999).
- Girard, M., Habel, A. & Chanel, C. New prospects for the development of a vaccine against human immunodeficiency virus type 1. An overview. C. R. Acad. Sci. III 322, 959–966 (1999).
- Kent, S. J. et al. Enhanced T-cell immunogenicity and protective efficacy of a human immunodefficiency virus type 1 vaccine regimen consisting of consecutive priming with DNA and boosting with recombinant fowlpox virus. J. Virol. 72, 10180–10188 (1998).
- J. Virol. 72, 10180–10188 (1998).

 100. Hel. Z. et al. Potentiation of simian immunodeficiency virus (SIV)-specific CD4* and CD8* T cell responses by a DNA-SIV and NYVAC-SIV prime/boost regimen.

 J. Immunol. 167, 7180–7191 (2001).

- 101. Mayr, A., Stickl, H., Muller, H. K., Danner, K. & Singer, H. [The smallpox vaccination strain MVA: marker, genetic structure, experience gained with the parenteral vaccination and behavior in organisms with a debilitated detence mechanism (author's transl.)]. Zentraitol. Bakteriol. [B] 167, 375–390 (1978).
- Stickl, H. et al. [MVA vaccination against smallpox: clinical tests with an attenuated live vaccinia virus strain (MVA) (author's transil). Disch. Med. Wochenschr. 99, 2388–2392 (1974).
- 103. Berouch, D. H. et al. Reduction of simien-human immunodeficiency virus 89. 6P viremia in rhesus monkeys by recombinant modified vaccinia virus Ankara. vaccination. J. Virol. 76, 5151–5158 (2001).
- Cromwell, M. A. et al. Induction of mucosal homing virusspecific CDB* T lymphocytes by attenuated simian inmunodeficiency virus. J. Viro. 74, 8782–8768 (2000).
 Gallimore, A. et al. Early suppression of SIV replication by
- 105. Gallimore, A. et al. Early suppression of SIV replication by CD8* nef-specific cytotoxic T cells in vaccinated macaques. Nature Med. 1, 1167–1173 (1995).
- 106. Fuller, D. H. et al. Gene gun-based nucleic acid immunization alone or in combination with recombinant vaccinia vectors suppresses virus burden in rhesus macaques challenged with a heterologous SIV. Immunol. Cell Biol. 75, 369-396 (1997).
- Allen, T. M. et al. Induction of AIDS virus-specific CTL activity in fresh, unstimulated peripheral blood lymphocytes from rhesus macaques veccinated with a DNA prime/modified vaccinia virus Ankara boost regimen. J. Immunol. 184, 4988–4978 (2000).
- Kaul, R. et al. HIV-1-specific mucosal CD8⁻ lymphocyte responses in the cervix of HIV-1-resistant prostitutes in Nairobi. J. Immunol. 164, 1602–1611 (2000).
- 109. Kaul, R. et al. Late seroconversion in HIV-resistant Nairobi prostitutes despite pre-existing HIV-specific CDS responses. J. Clin. Invest. 107, 341–349 (2001)
- responses. J. Clin. Invest. 107, 341–349 (2001). 110. Rosenberg, E. S., LaRosa, L., Flynn, T., Robbins, G. & Walker, B. D. Characterization of HIV-1-specific T-helper cells in acute and chronic infection. Immunol. Lett. 66, 89–93 (1999).
- Zhang, C., Comette, J. L., Berzofsky, J. A. & DeLisi, C. The organization of human leucocyte antigen class I epitopes in HIV genome products: implications for HIV evolution and vaccine design. Nanohe 16, 1291–1302 (1902).
- vaccine design. Vaccine 15, 1291–1302 (1997).

 112. Burrows, S. R. et al. T cell receptor repertoire for a viral epitope in furnans is diversified by tolerance to a background major histocompetibility complex antigen.

 J. Exp. Med. 182, 1703–1715 (1995).

 113. Klenerman, P. et al. Cytotoxic T-cell activity antagonized
- Klenerman, P. et al. Cytotoxic T-cell activity antagonized by naturally occurring HIV-1 Gag variants. Nature 369, 403–407 (1994).
- Reid, S. W. et al. Antagonist HIV-1 Gag peptides induce structural changes in HLA B8. J. Exp. Med. 184, 2279–2286 (1996).
- Klenerman, P. & Zinkernagel, R. M. Original antigenic sin impairs cytotoxic Tlymphocyte responses to viruses bearing variant epitopes. Nature 394, 482–485 (1998).

- Mortara, L. et al. Selection of virus variants and emergence of virus escape mutants after immunization with an epitope vaccine. J. Virol. 72, 1403–1410 (1998).
- Price, D. A. et al. Positive selection of HIV-1 cytotoxic
 T lymphocyte escape variants during primary infection.
 Proc. Natl Acad. Sci. USA 94, 1890-1895 (1997).
- Proc. Natl Acad. Sci. USA 94, 1890–1895 (1997).
 118. Kelleher, A. D. et al. Clustered mutations in HIV-1 gag are consistently required for escape from HLA-B27-restricted CTL responses. J. Exp. Med. 193, 375–386 (2001).
 119. Korber, B. et al. Evolutionary and immunological
- Korber, B. et al. Evolutionary and immunological implications of contemporary HIV-1 variation. Br. Med Bull. 58, 19–42 (2001).
- Goulder, P. J. et al. Evolution and transmission of stable CTL escape mutations in HIV infection. Nature 412, 334–338 (2001).
- 121. Hsu, S. C. et al. Protective cytotoxic T lymphocyte responses against pararmy.coviruses induced by epitopebased DNA vaccines: involvement of IPN-y. Int. Immunol. 10, 1441–1447 (1998)
- 10, 1441–1447 (1998).
 122. Kulkami, A. B., Connors, M., Firestone, C. Y., Morse, H. C.
 III & Murphy, B. R. The cytolytic activity of pulmonary CD8lymphocytes, induced by infection with a vaccinia virus recombinant expressing the M2 protein of respiratory syncytial virus (RSV), correlates with resistance to RSV infection in mice. J. Virol. 67, 1044–1049 (1993).
- infection in mice. J. Virol. 67, 1044–1049 (1993).

 123. Fu, T. M., Friedman, A., Ulmer, J. B., Liu, M. A. & Donnelly, J. J. Protective cellular immunity: cytotoxic T-lymphocyte responses against dominant and recessive epitopes of influenza virus nucleoprotein induced by DNA immunization. J. Virol. 71, 2715–2721 (1997).

 124. Rodriguez, F., Zheng, J. & Whitton, J. L. DNA
- 124. Rodríguez, F., Zhang, J. & Whitton, J. L. DNÁ immunization: ubiquitination of a viral protein enhances cytotoxic T-lymphocyte induction and antiviral protection but abrogates antibody induction. J. Virol. 71, 8497–8503 (1997).
- 125. Seth, A. et al. Immunization with a modified vaccinia virus expressing simian immunodeficiency virus (SIV) Gag.-Pol primes for an anemnestic Gag-specific cytotoxic T-lymphocyte response and is associated with reduction of viremia after SIV challenge. J. Virol. 74, 2502–2509 (2000).
- 126. Belyakov, I. M. et al. Mucosal AIDS vaccine reduces disease and viral load in gut reservoir and blood after mucosal infection of macaques. Nature Med. 7, 1320–1326 (2001).

Online links

DATABASES

The following terms in this article are linked online to: LocusLink: http://www.ncbi.nlm.nih.gov/LocusLink CCR5 | CD4 | CD8 | CXCR4 | IFN- α | IFN- γ | IL-2 | IL-12 | MIP-1 α

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Ethical issues for vaccines and immunization

Jeffrey B. Ulmer and Margaret A. Liu

Vaccination is the only type of medical intervention that has eliminated a disease successfully. However, both in countries with high immunization rates and in countries that are too impoverished to protect their citizens, many dilemmas and controversies surround immunization. This article describes some of the ethical issues involved, and presents some challenges and concepts for the global community.

Vaccines stand out as being among the most efficacious and cost-effective of global medical interventions¹ (BOX 1). Vaccines have saved millions of lives, prevented significant morbidity and suffering, and even eradicated a disease. This last accomplishment, the eradication of smallpox, highlights what can be achieved by vaccination. However, unfortunately, the inequalities in the distribution and use of vaccines are also striking. If vaccines